Reduction of 6. Lithium aluminum hydride was added to a solution of **6** (40 mg) in anhydrous THF. The mixture was refluxed for 26 h. Ether was added, and the solution was washed with 1 M H_2SO_4 , saturated NaHCO₃, and brine. After the mixture was dried over MgSO₄, the ether was removed in vacuo. Preparative TLC of the residue gave 7: 24 mg; EI mass spectrum, m/z (relataive intensity) $222 \ (\text{M}^+, 6)$, $207 \ (\text{4})$, $204 \ (26)$, 191 (7) , 189 (7), 179 (lo), 161 (49, 109 (100); 'H NMR, Table 11.

Reduction of 5. Zinc/acetic acid reduction of 1 (50 mg) gave, after the workup, 33 mg of a debrominated mixture (5-7), which was treated with LiAlH, in refluxing THF for 4.5 h. After the **usual** workup, preparative TLC afforded 7 (9 mg, 29%) and **8** (11 mg, 33%). For $\hat{\mathbf{8}}$: crystalline; CI mass spectrum [i-C₄H₁₀ + $(CH_2NH_2)_2$, m/z (relative intensity) 301 (\dot{M} + 61, 67), 283 (5), 265 (ll), 223 (loo), 205 *(85),* 121 (62); E1 mass spectrum, m/z (relative intensity) 222 (M - 18, 20), 207 (37), 204 *(5),* 189 (a), 179 (7), 161 (ll), 137 (loo), 121 (23), 109 (441, 95 (28), 81 (56), 43 (72); 'H NMR, Table 11.

Dehydration of 8: (+)-6-Selinene (9). A catalytic amount of p-toluenesulfonic acid was added to a benzene solution of **8** (11 mg). The mixture was refluxed for 45 min. Ether was added, and the organic phase was washed with saturated NaHC0, and brine. After the mixture was dried over MgSO₄, the solvent was removed in vacuo. Preparative TLC of the crude product (petroleum ether) gave 4 mg (43%) of $(+)$ - δ -selinene (9): $[\alpha]^{24.5}$ _D +195° (c 0.49, CHCl₃); EI mass spectrum, m/z (relative intensity) 204 (M', 93), 189 (loo), 161 (93), 147 (13), 133 (27), 119 (21), 105 (34), 95 (22), 91 (32), 81 (21), **55** (17), 40 (36); UV (EtOH) **A,,** 234 nm (sh), 248.5, 256 (sh); 'H NMR, Table 11.

Dehydration of 1 **with POCl,.** Phosphorus oxychloride (130 μ L in five portions, 10.5-fold excess) was added to a solution of 1 (47 mg) in pyridine (0.7 mL) during a total reaction time of 19 h at 25 °C and 12 h at 42 °C. When the reaction was complete, ether and ice were added. Pyridine was removed by two 3-mL washes of 3 M HCl. The ether solution, after being washed with saturated NaHCO₃ and brine, was dried over MgSO₄. After filtration, the ether was removed in vacuo. Preparative TLC (once with CH_2Cl_2) gave 38 mg of 10 (86%) and 3 mg of starting material. Yields were lower without a large excess of POCl₃. For 10: EI mass spectrum, m/z (relative intensity) 342/344 ($\overline{M}^+, 1$), 282/284 (49), 267/269 (3), 263 (2), 254/256 (4), 239/241 **(58),** 203 (72), 159 (51), 147 (26), 107 (31), 84 *(58),* 43 (100); **Et** (KBr) 2960, 2885,1720,1650,1460,1440,1390,1370,1270,1260,1245,1230, 1030, 885 cm-'; 'H NMR, Table 11.

Dehydrobromination of 10. DBU **(1,5-diazabicyclo[5.4.0]** undec-5-ene, 1.5 mL) was added to 42 mg of crude exo olefin 10, and the reaction mixture was heated at $105 °C$ for 4 days. The resulting *gum* was slowly dissolved with 3 M HCl(6 mL) and ether. After removal of the acid layer, which was back-washed with fresh ether, the total ether phase was washed with saturated NaHC0, and brine and dried over MgS0,. After filtration and vacuum removal of the solvent, preparative TLC of the crude (four times with 25% CH₂Cl₂/petroleum ether) gave minimal amounts of two less polar UV-active components and 11 as a faintly yellow oil: 23 mg (71%); E1 mass spectrum, *m/z* (relative intensity) 262 (M'), 247, 220,219,202 (37), 187 (25), 159 (loo), 145 (13), 131 (34), 117

(17), 105 (22), 81 (22),43 *(56);* 'H NMR, Table II; 13C *NMR,* Table I.

Dehydration of 1 with TsOH. A catalytic amount of ptoluenesulfonic acid was added to a solution of 1 (78 mg) in benzene (1 mL). The solution was refluxed on a steam bath for 4.5 h. The reaction was quenched with saturated NaHC0, and extracted with ether. The ether solution was washed with saturated NaHCO₃ and brine and dried over MgSO₄. After filtration, the ether was evaporated in vacuo. Preparative TLC (twice with petroleum ether) gave 12 (16 mg). Redevelopment of the remaining plate (once with CH_2Cl_2) gave 10 (38 mg) and 13 (1 mg). For $12^{.6}$ UV λ_{max} 236 nm (ϵ 4130), 248 (sh, 3710), 257 (sh, 2840), 299 (2180); 'H NMR, Table 11.

Hydrolysis of 10. A solution of 10 (38 mg) in benzene was added to a 9-fold excess of $KO-t-Bu$ in $Me₂SO$. The mixture was refluxed for 4.5 h. The products were extracted with water and ether. The ether solution was washed with saturated $NAHCO₃$ and brine and dried over MgSO₄. After filtration, the ether was removed in vacuo to give a quantitative yield of 13: E1 mass spectrum, m/z (relative intensity) 300/302 (M', l), 282/284 (13), 239/241 (lo), 221 (lo), 203 (39), 187/189 (21), 169 (la), 149 (27), 119 (43), 107 (96), 91 (43), 81 (54),69 (44), **55** (63), 41 (100); 'H NMR, Table 11.

Acetylation of 1. A solution of 1 (21 mg), pyridine (0.3 mL), acetic anhydride (0.3 mL), and **4-(dimethy1amino)pyridine** (17 mg) in CCl₄ (3 mL) was allowed to react for 6 days at 25° C. Ether was added to the mixture, and the organic phase was washed successively with dilute HCl, 10% NaHCO₃, and brine solutions. After drying over $MgSO_4$ and filtration, the ether was removed in vacuo. Preparative TLC of the crude product (33 mg; four times with CH_2Cl_2) gave 2 (1 mg, 4%), 14 (14 mg, 49%), and 15 (7 mg, 25%).

14: CI mass spectrum $(i-C_4H_{10})$ m/z (relative intensity) $487/489$ $(M + 1, 1), 427/429$ (3), $385/387$ (1), $343/345$ (84), $301/303$ (3), 283/285 (59), 263/265 (9), 239/241 (2), 227/229 (4), 221 (13), 205 (29), 203 (loo), 145 (36), 127 (19), 103 (9), 85 (25); IR (film) 2970, 2885,1765,1730,1715,1665,1460,1440,1390,1365,1340,1250, 1225,1205,1195,1110,1080,1035,1020,900,865,700 cm-'; 'H NMR, Table 11; 13C NMR, Table I.

15: CI mass spectrum $(i-C_4H_{10})$, m/z (relative intensity) $487/489$ (M + 1, 1), $427/429$ (2), $343/345$, (99), $301/303$ (3), 283/285 (49), 263/265 (12), 239/241 (2), 227/229 (4), 221 (ll), 205 (30), 203 (loo), 145 (37), 127 (181,103 (6), 85 (59); 'H NMR, Table 11; 13C NMR, Table I.

Acknowledgment. We thank Dr. R. J. Wells and the Roche Research Institute of Marine Pharmacology for providing the alga and the facilities **for** this investigation. We are also grateful to **Mr.** Frank Shea, Clark University, for providing some **of** the **'H** and 13C NMR spectra (NSF Equipment Grant DMR-8108697).

Registry No. 1, 82731-83-7; 2, 82731-84-8; 3, 71778-85-3; 4, 82769-15-1; 5,82731-85-9; 6,82731-86-0; 7,82795-50-4; 8,82731-87-1; **9,** 28624-28-4; 10, 82731-88-2; **11,** 82731-89-3; 12, 82731-90-6; 13, 82731-91-7; 14, 82740-45-2; 15, 82740-46-3.

Stereocontrolled Synthesis of 205 Steroidal Side Chain

Wiliam G. Dauben* and Todd Brookhart'

Department of Chemistry, University *of* California, Berkeley, California *94720*

Received December 29, *I981*

The diethylaluminum chloride catalyzed ene reaction between (20E)-3 β -acetoxy-5,17(20)-pregnadiene and methyl propiolate proceeds, stereospecifically, from the *a* face to yield methyl **(20S)-3@-acetoxychola-5,16,22-trienoate,** albeit at a slower rate than the *20R* formation from the *2* isomer. The triene was hydrogenated to yield the known methyl (20S)-3/3-acetoxycholanoate. Thus, the ene reaction can be used to prepare either *20R* or 20s steroidal side chains.

In the last decade, a wide variety of di- and sesterterpenes and steroids have been reported to have modified isooctyl (cholesterol-type side chains^{2,3} and the unit being attached to the polycyclic nucleus with *R* or *S* stereo-

chemistry in the isooctyl chain. The introduction of these side chains onto the polycyclic nucleus has been the subject of investigation by several research groups. Starting with a *(2)-* or an (E)-ethylidene side chain, both the R or S side chain have been uniquely introduced by alkylation of a $(\pi$ -allyl)palladium intermediate.⁴ In studies utilizing the Claisen-Carroll rearrangement for the stereocontrolled introduction of a side chain to a sterol, both the 20R and $20S$ isomers have been prepared.⁵ In a related study, however, employing the oxy-Cope rearrangement only the $20R$ isomer derived from the (Z) -17(20-pregnene derivative could be prepared.⁶ When the related (E) -17(20)-pregnene derivative was utilized, only reversion to the enones was found. It was suggested that the failure of this latter reaction to introduce the steroidal side chain presumably was due to the quasi-1,3-diaxial interaction between the 16-O⁻K⁺ and the 20-CH₃ groups.

Recently, it was reported from this laboratory,' and later by other workers,⁸ that (Z) -17(20)-pregnene derivatives on treatment with methyl propiolate and diethylaluminum chloride gave excellent yields of the (20R)-chola-16,22 dienic ester derivatives. In this reaction the well-known preference for attack on the α face of the C-17(20) double bonds⁹ and the highly ordered transition state of the ene rection set the stereochemistry in the natural 20R configuration. In view **of** the reported steric problem in the

oxy-Cope rearrangement using a (E) -17(20)-pregnene derivative, the ene reaction has been studied with the 17- (20)- \vec{E} isomer to evaluate the utility of the reaction sequence to prepare polycyclic materials with a steroidal side chain with an S configuration.

It has been found that treatment of 1^{10} with 1.2 equiv

(4) (a) Trost, B. M.; Bernstein, P. R.; Funfschilling, P. C. *J. Am. Chem.* SOC. **1979,101, 4378** and earlier papers. (b) Riediker, M.; Schwartz, J. Tetrhedron Lett. 1981, 4655 and earlier papers.
(5) Tanabe, M.; Hayashi, K. J. Am. Chem. Soc. 1980, 102, 862.
(6) Koreeda, M.; Tanaka, Y.; Schwartz, A. J. Org. Chem. 1980, 45,

1172.

(7) Dauben, W. **G.;** Brookhart, T. *J. Am. Chem.* SOC. **1981,103, 237. (8)** Batcho, **A.** D.; Berger, D. E.; Uskokovic, M. R.; Snider, B. B. *J. Am. Chem.* SOC. **1981,103,1293.** Batcho, **A.** D.; Berger D. E.; Davoust, S. *G.;*

Wovkulich, P. M.; Uskokovic, M. R. *Helu. Chim. Acta* **1981, 64, 1682. (9)** (a) Krubiner, A. M.; Oliveto, E. **P.** J. *Org. Chem.* **1966,31, 24.** (b) Krubiner, A. M.; Oliveto, E. P. *Zbid.* **1968,** *33,* **1715.**

of methyl propiolate with 2.0 equiv of diethylaluminum chloride in benzene at room temperature for **5** days gave a 32 mixture of starting material 1 and product **2.** In order to establish the structure and stereochemistry at C-20 in triene **2,** the material was hydrogenated to give the known hexahydro derivative **3.** The physical and spectral properties of **3** were virtually identical with those reported earlier." No trace of the 20R stereoisomer of **2** or **3** could be detected.

Thus, the ene reaction with the E olefin proceeds at least an order of magnitude less rapidly than with the *2* isomer, but the reaction still proceeds in a stereospecific manner. The nature of this rate retardation is not clear. This study broadens the present methodology so as to permit stereospecific synthesis of both $20R$ and $20S$ steroidal side chains.^{11a}

Experimental Section

Methyl $(20S, 22E)$ -3 β -Acetoxychola-5,16,22-trienate (2) . To a solution of **490** mg of olefin **1** (prepared by the method of Butenandt et **al.'O)** and **0.155** mL **(1.7** equiv) of methyl propiolate in **10** mL of dry benzene under an atmosphere of nitrogen was added, dropwise, **1.79** mL **(2.0** equiv) of a **25%** solution of diethylaluminum chloride in toluene. The reaction mixture was stirred at room temperature for **5** days, poured into **50** mL of a **5%** aqueous sodium bicarbonate solution, and extracted with ether. The organic layer was washed with an aqueous saturated salt solution and dried (MgSO₄) and the solvent removed under reduced pressure to afford a mixture of starting olefin **1** and product **2.** The mixture was chromatographed on silica with a **10%** ether-hexane mixture **as** the eluent. The less polar fraction contained **297** mg of **1,** while the more polar fraction contained **226** mg of **2 (94%** based upon recovered starting material). The product was recrystallized from methanol to produce a white solid mp **121-122** "C; **[c~]~-66.9"** (CHCl,); IR (CHC13) **1720,1650,1030, 980** cm-'; 'H NMR (CDClJ 6 **6.92** (dd, **1,** *J* = **8, 16** Hz), **5.80** (dd, **1, 1 1 1.5, 16 Hz**), **5.43** (m, 2), 3.75 (s, 3), 3.04 (qt, 1, **J** = 7, 8 **Hz**), **1. J** = 1.5, 16 **Hz**), **5.43** (m, 2), 3.75 (s, 3), 3.04 (qt, 1, **J** = 7, 8 **Hz**), 1.22 (d, 3, $J = 7$ Hz), 1.08 (s, 3), 0.81 (s, 3); ¹³C NMR (CDCl₃) δ **170.3, 167.2, 156.2, 153.4, 139.7, 123.8, 122.3, 119.0,73.7, 56.8,51.3,**

⁽¹⁾ NIH Postdoctoral Fellow, **1979-1981. (2)** (a) Nakanishi, **K.; Goto,** T.; Ito, S.; Natori, S.; Nozoe, S. "Natural Producta Chemistry"; Academic Press: New York, **1974;** Vol. I. (b) Nes, W. R.; McKean, M. L. 'Biochemistry **of** Steroids and Other Isoprenoids"; University Park Press: Baltimore, MD, **1977.**

⁽³⁾ (a) Hikino, H.; Hikino, Y. *Fortschr. Chem. Org. Naturst.* **1970,28, 256.** (b) Minale, L.; Sodano, *G.* In "Marine Natural Producta Chemistry"; Faulkner, D. **J.;** Fenical, W. H., **Eds.;** Plenum Press: New York, **1977.** (c) Norman, A. W. 'Vitamin D, The Calcium Homeostatic Steroid Hormone"; Academic Press: New York, **1979.** (d) DeLuca, **H.** F. 'Vitamin D Metabolism and Function"; Springer-Verlag: New York, **1979.**

⁽¹⁰⁾ Butenandt, A.; Schmidt-Thome, J.; Paul, H. *Chem. Ber.* **1939,72,** 112. The compound was recrystallized from ethanol to constant melting point [138–140 °C (lit.¹⁰ mp 140 °C). For this *E* isomer the ¹³C NMR spectrum showed four singlet absorptions for the olefinic trigonal carbon atoms at 6 **110.3, 122.5, 139.7,** and **152.3** and the 'H NMR spectrum showed olefinic proton'absorption at **6 5.05 (1,** m) and **5.39 (1,** d). For the earlier reported *Z* isomer,⁷ the ¹³C NMR values were δ 113.3, 127.1, 139.3, and 149.5 and the ¹H NMR values were δ 5.13 (1, m) and 5.38 (1, d).

⁽¹¹⁾ Vanderah, D. J.; Djerassi, C. *J. Org. Chem.* **1978, 43, 1442.** (a) Note Added in Proof: Since submission ion of this manuscript, M. M. Midland and Y. C. Kwon, *Tetrahedron Lett.* **1982, 2077,** have reported the synthesis of **20R-** and 20s-cholesterol derivatives by their hydroboration sequence.

50.3, 47.0, 38.0, 36.8, 36.6, 35.0, 34.8; 31.3, 31.1, 30.3, 27.6, 21.3, 20.6,19.8,19.1,16.1; mass **spectrum (70** eV), *m/e* **366 (9.20** parent - **CH3CO2H,** base), **351 (2.89), 253 (2.78).** Anal. Calcd for **C27H3804: C, 76.02; H, 8.98.** Found: **C, 75.86; H, 8.95.**

Methyl (205)-38-Acetoxy-5a-cholanoate (3). A solution containing **30** mg of **2** and **5** mg of **5%** PtO, in **12** mL of ethyl acetate was stirred under an atmosphere of hydrogen at ambient pressure for 12 h. The reaction mixture was filtered, the solvent was removed under reduced pressure, and the solid was recrystallized twice from methanol to produce white crystals, mp **134-135 "C** [lit." mp **136.0-137.5 "C].** The 'H **NMR** spectrum showed the C-21 absorption at δ 0.83 $(J = 6$ Hz) [lit.¹¹ C-21, δ 0.83 (12) The value for C-21 in the 20R series is δ 0.92 $(J = 6$ Hz).

 $(J = 6 \text{ Hz})$ ¹² The mass spectrum was identical with the literature values.¹¹

Acknowledgment. This work was supported by Grant No. AM 00709, National Institute of Arthritis, Diabetes, and Digestive and Kidney Diseases. We are greatly indebted to Dr. Jaan Pesti for his assistance.

Registry No. 1, 29842-93-1; 2, 82660-25-1; 3, 1178-02-5; methyl propiolate, 922-67-8.

Mass Spectrometry of Nucleic Acid Constituents. Trimethylsilyl Derivatives of Nucleosides^{1,2}

Henrianna Pang, Karl H. Schram, David L. Smith, S. P. Gupta, Leroy B. Townsend, and James A. McCloskey*

Departments of Medicinal Chemistry and Biochemistry, University of Utah, Salt Lake City, Utah 84112

Received May 4, 1982

Trimethylsilylation of nucleosides provides derivatives which are thermally volatile and whose electron-ionization mass spectra are useful for structural characterization and for determination of chemically or biologically incorporated stable **isotopes.** The major reaction pathways and mechanisms of fragmentation of silylated nucleosides have been studied, on the basis of the mass spectra of a structural variety of nucleoside analogues and of uridine selectively labeled with deuterium $(C-2', C-3', C-5', Si(CD₃)₃$ and oxygen-18 $(O², O⁴, O-2', O-3', O-5')$. Formation of most of the major base-containing ions, which are even-electron species, involving rearrangement of hydrogen from the sugar skeleton to the ionized base. The site selectivity of some of the rearrangement processes indicates that **base-H-2'** interactions are relatively important and that in those cases significant opening qf the ribose ring does not occur prior to hydrogen abstraction by the base. The relative abundance of sugar **H** ions resulting from transfer of **H-2'** to the base was found to be greater in derivatives of 6-ribofuranosides compared with that for the corresponding α anomers, reflecting differences in steric accessibility of H-2' to the base and providing a means of distinguishing α and β anomers. The determination of the site and extent of isotopic substitution in the sugar is better measured from fragment ions which contain the base plus portions of the sugar than from sugar ions which do not contain the base. This is a consequence of the multiple pathways of formation of most sugar-derived ions.

The structure elucidation of new nucleosides isolated from natural sources is often confounded by the combination of structural complexity and severe limitations in sample quantity. These problems are particularly prevalent in the case of transfer RNA, where modified nucleosides often occur at a rate of one residue per tRNA molecule, and the isolation of more than a few micrograms from grams or more of starting material is a demanding task.3 Mass spectrometry has thus evolved as an important technique for the characterization of nucleosides, 4 largely **as** a consequence of high sensitivity compared with other methods.

Direct vaporization techniques $5-13$ such as field desorp-

- **(2) A preliminary account of a portion of** this **work has been presented:**
- Hattox, S. E.; Schram, K. H.; McCloskey, J. A. 23rd Annual Conference
on Mass Spectrometry and Allied Topics, Houston May 1975, pp 435–436.
(3) McCloskey, J. A.; Nishimura, S. Acc. Chem. Res. 1977, 10, 403–410.
(4) Hignite **First Supplementary Volume"; Waller, G. R., Dermer, 0. C., Eds; Wiley:**
- **New York, 1980; Chapter 5. (5) Ohashi, M.; Tsujimoto, K.; Yasuda, A. Chem.** *Lett.* **1976 439-440. (6) Hunt, D. F.; Shabanowitz, J.; Botz, F. K. Anal.** *Chem.* **1977, 49,**

1160-1163.

- **(7) Posthumus, M. A.; Kistemaker, P.** *G.;* **Meuzelaar, H. L. C.; Ten (8) Lai, S.-T. F.; Evans, C. A., Jr. Biomed. Mass Spectrom. 1979, 6, Noever de Brauw, M. C. Anal.** *Chem.* **1978,50,985-991.**
- **10-14.**

tion¹⁴ or fast atom bombardment^{15,16} can be useful for nucleosides that cannot be volatilized thermally. However, the lower abundances of information-bearing fragment ions produced make these methods generally less desirable **as** a single approach compared with microscale derivatization and electron ionization. Of the various derivatization procedures which have been used for mass spectrometry of nucleosides, 4.17 the most widely used has been trimethylsilylation,¹⁸ which has been employed with partic-

- **(9) Cotter, R. J.; Fenselau, C. Biomed.** *Mass Spectrom.* **1979,** *6,* **287-293.**
- **(10) Esmans, E. L.; Freyne, E. J.; Vanbroeckhoven,** J. **H.; Alderweir eldt, F. C. Biomed.** *Muss* **Spectrom. 1980, 7, 377-380.**
- **(11) Le Beyec, Y.; Della Negra,** *S.;* **Deprun, C.; Vigny, P.; Ginot, Y. M. Reu.** *Phys.* **Appl. 1980, 15, 1631-1637.**
- (12) Blakley, C. R.; Carmody, J. J.; Vestal, M. L. *J. Am. Chem. Soc.* **1980,102, 5931-5933.**
- **(13) McNeal, C. J.; Narang, S. A.; Macfarlane, R. D.; Hsiung, H. M.; Brousseau, R. Proc. Natl. Acad. Sci. U.S.A. 1980, 77,735-739.**
- **(14) Schulten, H.-R.; Beckey, H. D.** *Org.* **Mass Spectrom. 1973, 7, 861-867.**
- **(15) Barber, M.; Bordoli, R.** *S.;* **Sedgwick, R. D. Nature** *(London)* **1981, 293, 270-275.**
- **(16) Williams, D. H.; Bradley, C.; Bojesen, G.; Santikarn,** S.; **Taylor, L. C. E.** *J.* **Am.** *Chem.* **SOC. 1981,103, 57OC-5704.**
- **(17) Hignite, C. In "Biochemical Applications of Mass Spectrometry"; Waller, G. R., Ed.; Wiley: New York, 1972; Chapter 5.**
- **(18) McCloskey, J. A.; Lawson, A.** M.; **Tsuboyama, K.; Krueger, P. M.; Stillwell, R. N.** *J.* **Am.** *Chem.* **SOC. 1968,** *90,* **4182-4184.**

⁽¹⁾ Taken in part from the Ph.D. thesis of H.P., University of Utah, 1982.